

## Protocol for Western Blotting

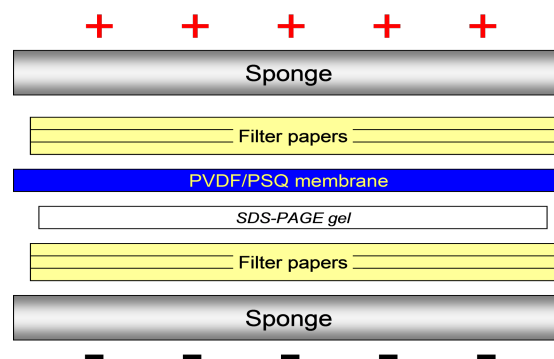
**RED TEXT** indicates important optimized settings

### SDS-PAGE separation

1. **Make separation gel (SDS-page gel in Tris-Glycine system) with a gradient from 8-14% .**
2. Prepare 30 µg **HEK-293 lysate** with **RIPA buffer** by mixing 4X SDS sample buffer with lysate sample according to the protein concentration measured by Bradford or BCA protein assay.
3. Heat at 95-100°C for 5 min. Set up the electrophoresis apparatus with SDS-PAGE gel.
4. Load samples and protein markers onto the gel. Set 80 V to run the stacking gel, increase to 120 V for the separation gel till the end.

### Electrotransfer

5. Soak **PVDF membrane with pore size 0.45 µm** in methanol for 30 sec and then in the transfer buffer. Soak the filter papers and sponges in the transfer buffer as well.
6. Sequentially assemble the transfer sandwich according to the illustration and make sure no bubbles are trapped. Apply **Wet transfer systems at 180mA for 90 min with Wet transfer buffer.**



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## Immunoblotting

7. After transferring, wash the membrane twice with distilled water.
8. Block membrane with 1X TBST containing 5% nonfat dry milk with agitation at room temperature for 1 h.
9. Dilute primary antibody (**18420-1-AP, P62/SQSTM1 Rabbit PolyAb**) **1:1000** in blocking solution. Incubate membrane with the primary antibody with agitation at **room temperature for 1.5 hours**. Wash membrane 3 times with 1X TBST for 10 min each.
10. Incubate the membrane with the HRP-conjugated secondary antibody diluted at **1:5000** in blocking solution **at room temperature for 1 h**. Wash membrane 3 times with 1X TBST for 10 min each.

## Signal Detection (Chemiluminescence system)

11. Prepare ECL substrate according to the manufacturer's instructions.
12. Incubate the membrane with a substrate for 1-5 min.
13. Exposure autoradiography film in the dark or try a chemiluminescence imaging system. Check a few different exposure times to obtain the optimal image for quantification (30-300 sec).
14. Remember to mark the protein MW markers on the film.

## Solutions

<b>RIPA buffer</b>	<b>1000 ml</b>
50 mM Tris-HCl, pH 7.4 (1M stock)	50 ml
150 mM NaCl	8.76 g
1% Triton X-100 or NP-40	10 ml
0.5% Sodium deoxycholate	5 g
0.1 % SDS	1 g
1 mM EDTA (0.5 M stock)	2 ml
Add ddH <sub>2</sub> O to 1000 ml	
Add PMSF to 1mM and protease inhibitors (1 mM Sodium Orthovanadate) freshly before use.	

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<b><u>4X SDS sample buffer</u></b>	<b>50 ml</b>	<b><u>1X TBST</u></b>	<b>1000 ml</b>
250 mM Tris-HCl (pH 7.0) (1M stock)	12.5 ml	20 mM Tris-base	2.4 g
40% glycerol	20 ml	150 mM NaCl	8.7 g
5% SDS	2.5 g	0.2% Tween-20	2 ml
0.02% Bromophenol Blue	10 mg	Adjust pH to 7.6	
5% β-mercaptoethanol	2.5 ml	Add ddH <sub>2</sub> O to 1000ml	
Add ddH <sub>2</sub> O to 50ml, aliquot and store at -20°C			

<b><u>1X Running buffer</u></b>	<b>1000 ml</b>	<b><u>Semi-dry transfer buffer</u></b>	<b>1000 ml</b>
12.5 mM Tris-base	1.51 g	48 mM Tris-base	5.81 g
100 mM Glycine	7.5 g	39 mM Glycine	2.93 g
0.05% SDS	0.5 g	0.0375% SDS	0.375 g
Add ddH <sub>2</sub> O to 1000 ml		20% Methanol	200 ml
		Add ddH <sub>2</sub> O to 1000 ml	

<b><u>Wet transfer buffer</u></b>	<b>1000 ml</b>	<b><u>Tricine gel running buffer</u></b>	<b>1000 ml</b>
25 mM Tris-base	3.03 g	100 mM Tris-base	12.1 g
192 mM Glycine	14.4 g	100 mM Tricine	17.9 g
20% Methanol	200 ml	0.1% SDS	1 g
Add ddH <sub>2</sub> O to 1000 ml		Add ddH <sub>2</sub> O to 1000ml	

## Related Products

Product Name	Catalog No.	Size	Application
HRP-conjugated AffiniPure Goat Anti-Mouse Ig(G+L)	SA00001-1	100 µl	ELISA; WB; IHC/ICC
HRP-conjugated AffiniPure Goat Anti-Rabbit Ig(G+L)	SA00001-2	100 µl	ELISA; WB; IHC/ICC

*ELISA: Enzyme-linked immunosorbent assay; WB: Western Blotting; IHC: Immunohistochemistry; ICC: Immunocytochemistry;*

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