

Premo™ Autophagy Tandem Sensor RFP-GFP-LC3B Kit

Catalog no. P36239

Table 1 Contents and storage

Material	Amount	Concentration	Storage*
RFP-GFP-LC3B (Component A)	1 mL	~1 × 10 ⁸ viral particles/mL	<ul style="list-style-type: none"> • 2–8°C • Protect from light • DO NOT FREEZE
Chloroquine diphosphate (Component B)	1 mL	30 mM aqueous solution	<ul style="list-style-type: none"> • ≤25°C • Protect from light
Approximate fluorescence excitation/emission maxima: Emerald GFP: 488/509 in nm; TagRFP: 555/584 in nm.			
* When stored as directed, this kit is stable for at least 6 months.			

Introduction

Autophagy describes the segregation and delivery of cytoplasmic cargo, including proteins and organelles, for degradation by hydrolytic enzymes through the lysosomal machinery. Although first described in 1963, it has only been in the past decade that this pathway has been the subject of intense research to gain further insight into the role basal autophagy plays in cell homeostasis and development (Figure 1, page 2). Efforts are also directed to further elucidate the role of induced autophagy as a cell survival response to stress, microbial infection, and disease (e.g., neurodegeneration, cancer).^{1–3}

Single FP (fluorescent protein) chimeras such as the Premo™ Autophagy Sensor LC3B-GFP (Cat. no. P36235) or LC3B-RFP (Cat. no. P36236) are useful for monitoring autophagosome formation, especially when combined with other FP chimeras or fluorescent reagents. However, the use of a tandem FP construct that allows an enhanced dissection of the maturation of the autophagosome to the autolysosome has recently been described.^{4,5} By combining an acid-sensitive GFP (i.e., Emerald GFP) with an acid-insensitive RFP (i.e., TagRFP), the change from an autophagosome (neutral pH) to the autolysosome (with an acidic pH) can be visualized by imaging the specific loss of the GFP fluorescence upon acidification of the autophagosome following lysosomal fusion. Upon induction of autophagy, the Premo™ Tandem Autophagy Sensor labels the punctate autophagosomes; these structures are positive for both GFP and RFP. Once the lysosome has fused, the pH drops, which quenches the GFP, making autolysosomes appear red (Figure 2, page 2, and Figure 3, page 3). Combining Premo™ Autophagy Tandem Sensor with the far-red emitting LysoTracker® Deep Red allows for a three-color analysis of the autophagosomal/autolysosomal/lysosomal dynamics.

For Research Use Only. Not for use in diagnostic procedures.

The Premo™ Autophagy Tandem Sensor combines the selectivity of an LC3B-fluorescent protein (FP) chimera with the transduction efficiency of the BacMam 2.0 technology. BacMam reagents (insect **B**aculovirus with a **M**ammalian promoter) are safe to handle (Biosafety Level 1) because they are non-replicating in mammalian cells. They are also non-cytotoxic and ready-to-use. Unlike expression vectors, BacMam reagents enable titratable and reproducible expression and offer high co-transduction efficiency. Recent improvements made to the BacMam system, BacMam 2.0, enable efficient transduction in a wider variety of cells, including neurons and neural stem cells (NSCs).

Each Premo™ Autophagy Sensor Kit includes chloroquine diphosphate for artificially generating autophagosomes. Following treatment with chloroquine diphosphate, normal autophagic flux is disrupted and autophagosomes accumulate as a result of the increased lysosomal pH that inhibits lysosomal fusion with the autophagosomes.

Figure 1 Schematic depiction of the autophagy pathway in a eukaryotic cell. The first step involves the formation and elongation of the isolation membranes or phagophore. The second step entails the expansion and sequestering of the cytoplasm and formation of the double-membrane autophagosome and includes the association of the cytosolic LC3B protein. Fusion of lysosomes with autophagosome to generate the autolysosome is the penultimate step. In the fourth and final phase, the cargo is degraded.

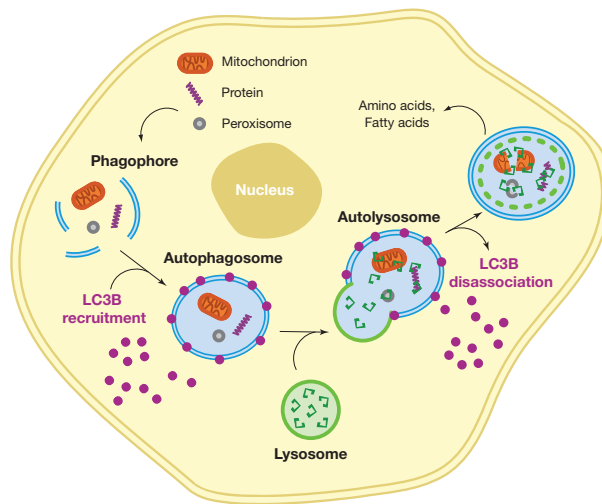


Figure 2 The use of the Premo™ Autophagy Tandem Sensor RFP-GFP-LC3B allows for an enhanced dissection of the maturation of the autophagosome to the autolysosome. By combining an acid-sensitive GFP with an acid-insensitive RFP, the change from autophagosome (neutral pH) to autolysosome (with an acidic pH) can be visualized by imaging the specific loss of the GFP fluorescence, leaving only red fluorescence.

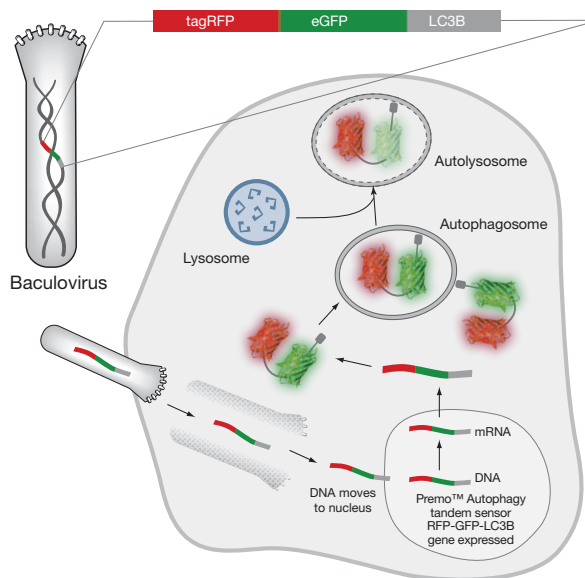
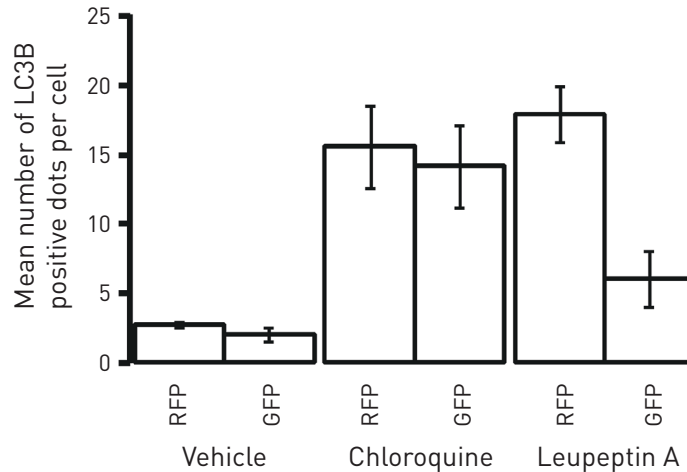


Figure 3 Premo™ Autophagy Tandem Sensor RFP-GFP-LC3B can be used to discriminate between acidic and neutral LC3B-positive vesicles. HeLa cells were transduced with 40 particles per cell of the Premo™ Autophagy Tandem Sensor and cultured for 24 hours. Cells were then incubated in either Vehicle, 90 μM Chloroquine or 200 μM Leupeptin A for an additional 24 hours. Cells were imaged using standard FITC/TRITC filter sets. Chloroquine blocks autophagy through neutralization of lysosomal pH; as a result, fluorescence is seen from both GFP and RFP, reflected in a significantly higher number of GFP and RFP positive spots compared to vehicle treated cells. Leupeptin A blocks autophagy without changing lysosomal pH; consequently, GFP fluorescence is lost in the acidic environment of the autolysosome whereas TagRFP retains its fluorescence. Therefore, a significant increase in the number of RFP positive spots with a decrease in the number of GFP positive spots is observed in Leupeptin A-treated cells.



Before Starting

Materials Required but Not Provided

- Cell culture medium

Guidelines for Working with BacMam Reagents

- The following protocol is based on a 2 mL labeling volume and ~40,000 cells plated in a 35-mm dish or 1 well of a 6-well culture plate, with a PPC (particles per cell) of 30.
- For applications that require a larger number of cells, such as high-content screening (HCS), we recommend transducing the cells in a 10-cm dish or a T-75 flask and increasing the labeling volume to 10 mL with a proportionate increase in the volume of the virus. Following an overnight incubation under normal growth conditions, trypsinize and count cells for distribution to appropriate plates at cell number desired.
- For optimal results you may alter the PPC, cell density, temperature, and incubation time.
- We recommend transducing cells at a confluency of about 70% for best results.
- For first time users of BacMam reagents, we recommend using the exceptionally well-transduced cells like U-2 OS (ATCC CCL-11226).

Experimental Protocols

Protocol The following protocol was optimized using cells that have been plated. You can also treat the cells in suspension prior to plating.

- 1.1 Plate the cells at the desired density and allow them sufficient time to adhere. Premo™ Autophagy Sensors work best when used on low-passage-number cells.
- 1.2 Calculate the volume of the tandem LC3B-FP (Component A) using the equation below:

$$\text{mL of tandem LC3B-FP} = \frac{(\text{number of cells}) (\text{PPC})}{(1 \times 10^8 \text{ Premo}^{\text{TM}} \text{ particles/mL})}$$

where the number of cells is the estimated total number of cells at the time of cell labeling, PPC (particles per cell) is the number of viral particles per cell, and 1×10^8 is the number of viral particles per mL of the reagent.

For example, to label 40,000 cells with a PPC of 30,

$$\text{mL of tandem LC3B-FP} = \frac{(40,000) (30)}{(1 \times 10^8 \text{ Premo}^{\text{TM}} \text{ particles/mL})} = 0.012 \text{ mL (12 } \mu\text{L)}$$

- 1.3 Mix each LC3B reagent several times by inversion to ensure a homogenous solution. **Do not vortex the LC3B reagent.**
- 1.4 Add the LC3B reagent directly to the cells in complete cell medium and mix gently.
- 1.5 Incubate the cells overnight (≥ 16 hours).
- 1.6 *Optional:* Treat the control cells with 30–100 μM chloroquine (Component B) for 12–16 hours.

Note: For best results, allow cells 48 hours post-transduction for expression levels to equilibrate.^{6,7}
- 1.7 Image and analyze the cells using the appropriate instrument filter sets for GFP and RFP detection. Autophagosomes are typically located in the perinuclear region.

Notes

- A. Premo™ Autophagy Tandem Sensors were designed for use in live-cell imaging of autophagy. The cell-permeant nucleic acid stains Hoechst 33342 and HCS NuclearMask™ Blue stains are spectrally compatible with the Premo™ Autophagy Tandem Sensor fluorescence. Should you prefer fixed cell analysis, the fluorescence from GFP and RFP has been demonstrated to be resistant to fixation with 4% formaldehyde and permeabilization with 0.1% Triton® X-100. Fixation and permeabilization enables processing of labeled cells with antibodies to other cellular targets. We recommend fixed cell format for large sample sizes, such as for HCS. Note that the fixation treatment may need to be optimized, and that certain treatments may change the pH and thus alter the fluorescence intensity of either fluorescent protein.

- B.** Be aware of bleed-through from one fluorescent protein into the detection channel of another,⁶ especially with the excitation of TagRFP by GFP filter sets and the differential brightness of GFP versus TagRFP.
- C.** GFP and TagRFP have differential photostabilities. Therefore, artifacts caused by photobleaching during time-lapse imaging should be taken into consideration.
- D.** Drug treatments may be used to calibrate the system. Chloroquine (Component B) is an excellent drug to block autophagic flux, which causes the accumulation of GFP- and TagRFP-positive vesicles. These protocols allow for the extent of bleed-through and bleaching to be examined. Note that Chloroquine inhibits autophagy by neutralizing lysosomal pH. The hydrolases in the lysosome require an acidic environment to function; even mild alkalinization inhibits these enzymes. Conditions that inhibit the enzymes may not be sufficient to alkalinize the autolysosomes sufficiently to rescue GFP fluorescence. For this reason, chloroquine should be used at relatively high concentrations (>90 μ M) for at least 12 hours. Leupeptin A or other drugs such as an E64D/Pepstatin A cocktail inhibit lysosomal enzymes without affecting pH and can be used to block autophagy, resulting in an accumulation of autolysosomes that maintain their acidity, thereby quenching GFP fluorescence. We recommend 200 μ M Leupeptin A treatment for 18 hours. Under these conditions, cells with predominantly RFP-positive and GFP-negative vesicles are seen. Earle's Balanced Salt Solution (EBSS) nutrient deprivation is also a useful control that allows for the progression from diffuse GFP/RFP fluorescence to GFP/RFP-positive autophagosomes, followed by RFP-positive autolysosomes.

References

1. Molecular Cell Biology 8, 931 (2007); 2. Genes & Development 21, 2861 (2007); 3. Drug Discovery 6, 304 (2007); 4. J Cell Biol 171, 603 (2005); 5. FEBS Letts 512, 19 (2002); 6. J Biol Chem 282, 24131 (2007); 7. Meth Enz 452, 25 (2009); 8. Meth Enz 452, 181 (2009); 9. Autophagy 8, 445 (2012).

Product List Current prices may be obtained from our website or from our Customer Service Department.

Cat. no.	Product Name	Unit Size
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Related Products		
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P36236	Premo™ Autophagy Sensor LC3B-RFP *BacMam 2.0*	1 kit
P36240	Premo™ Autophagy Sensor GFP-p62	1 kit
P36241	Premo™ Autophagy Sensor RFP-p62	1 kit
C10596	CellLight™ Lysosomes-GFP *BacMam 2.0*	1 mL
C10597	CellLight™ Lysosomes-RFP *BacMam 2.0*	1 mL
C10600	CellLight™ Mitochondria-GFP *BacMam 2.0*	1 mL
C10601	CellLight™ Mitochondria-RFP *BacMam 2.0*	1 mL
H3570	Hoechst 33342, trihydrochloride, trihydrate *10 mg/mL solution in water*	10 mL
H10325	HCS NuclearMask™ Blue	65 μ L
L7528	LysoTracker® Red DND-99 *1 mM solution in DMSO* *special packaging*	10 \times 50 μ L
L7535	LysoTracker® Red DND-26 *1 mM solution in DMSO* *special packaging*	10 \times 50 μ L
L10382	LC3B Antibody Kit for Autophagy *rabbit polyclonal LC3B* *includes autophagosome inducer*	1 kit
M7510	MitoTracker® Orange CMTMRos *special packaging*	10 \times 50 μ L
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M22423	MitoTracker® Deep Red *special packaging*	10 \times 50 μ L

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