



## Recommended Reaction Conditions

- 1X FastDigest Buffer or 1X FastDigest Green Buffer.
- Incubation at 37°C.
- 1 µL of FastDigest Eco52I is formulated to digest up to:
  - 1 µg of lambda DNA in 5 min.
  - 1 µg of plasmid DNA in 20 min.
  - 0.2 µg of PCR product in 20 min.
  - 1 µg of genomic DNA in 5 min, or 5 µg of genomic DNA in 30 min.

**Thermal Inactivation:** Incubation at 65°C for 5 min.

## Methylation Effects on Digestion

Dam: never overlaps – no effect.

Dcm: never overlaps – no effect.

CpG: completely overlaps – blocked.

EcoKI: never overlaps – no effect.

EcoBI: never overlaps – no effect.

## Compatible Ends

Check [www.thermoscientific.com/research](http://www.thermoscientific.com/research) for the list of restriction enzymes producing compatible ends.

## Number of Recognition Sites in DNA

λ	ΦX174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
2	0	1	0	0	0	0

## CERTIFICATE OF ANALYSIS

### Functional Activity Test

1 µg of lambda DNA-Eco81I fragments was completely digested with 1 µL of the enzyme in 5 minutes at 37°C in 20 µL of reaction mixture.

### Ligation and Recleavage (L/R) Assay

The ligation and recleavage assay was replaced with LO test after validating experiments showed LO test ability to trace nuclease and phosphatase activities with sensitivity that is higher than L/R by a factor of 100.

### Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or double-stranded oligonucleotides occurred during incubation with 1 µL of FastDigest Eco52I for 1 hour.

### Prolonged Incubation / Star Activity Assay

No detectable degradation of 1 µg of lambda DNA due to nuclease contamination or star activity occurred during incubation with 1 µL of FastDigest Eco52I for 16 hours.

### Blue/White (B/W) Cloning Assay

The B/W assay was replaced with LO test after validating experiments showed LO test ability to detect nuclease and phosphatase activities with sensitivity that equals to that of B/W test.

Quality authorized by:

 Jurgita Zilinskiene

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## Protocol for Fast Digestion of Different DNA

1. Combine the following reaction components at room temperature in the order indicated:

	Plasmid DNA	PCR product	Genomic DNA
Water, nuclease-free (#R0581)	15 $\mu$ L	17 $\mu$ L	30 $\mu$ L
10X FastDigest or 10X FastDigest Green Buffer	2 $\mu$ L	2 $\mu$ L	5 $\mu$ L
DNA	2 $\mu$ L (up to 1 $\mu$ g)	10 $\mu$ L (~0.2 $\mu$ g)	10 $\mu$ L (5 $\mu$ g)
FastDigest enzyme	1 $\mu$ L	1 $\mu$ L	5 $\mu$ L
Total volume:	20 $\mu$ L	30 $\mu$ L	50 $\mu$ L

2. Mix gently and spin down.

3. Incubate at 37°C in a heat block or water thermostat for 5 min (genomic DNA), or for 20 min (PCR product and plasmid DNA).

Optional: Inactivate the enzyme by heating for 5 min at 65°C.

4. If the FastDigest Green Buffer was used in the reaction, load an aliquot of the reaction mixture directly on a gel.

**Note:** The FastDigest Green Buffer can be used as an electrophoresis loading buffer for any DNA sample at a final 1X concentration. Higher concentrations of FastDigest Green Buffer in the sample supply excess salt concentration which may alter DNA mobility.

### Double and Multiple Digestion of DNA

- The combined volume of the enzymes in the reaction mixture should not exceed 1/10 of the total reaction volume.
- Use 1  $\mu$ L of each enzyme and scale up the reaction conditions appropriately.
- If the enzymes require different reaction temperatures, start with the enzyme that requires a lower temperature, then add the second enzyme and incubate at the higher temperature.

### Scaling up Plasmid DNA Digestion Reaction

DNA	1 $\mu$ g	2 $\mu$ g	3 $\mu$ g	4 $\mu$ g	5 $\mu$ g
FastDigest enzyme	1 $\mu$ L	2 $\mu$ L	3 $\mu$ L	4 $\mu$ L	5 $\mu$ L
10X FastDigest or 10X FastDigest Green Buffer	2 $\mu$ L	2 $\mu$ L	3 $\mu$ L	4 $\mu$ L	5 $\mu$ L
Total volume:	20 $\mu$ L	20 $\mu$ L	30 $\mu$ L	40 $\mu$ L	50 $\mu$ L

**Note:** Increase the incubation time by 3-5 min if the total reaction volume exceeds 20  $\mu$ L. Use water thermostat, air thermostats are not recommended due to the slow transfer of heat to the reaction mixture.

## Recommendations for PCR product digestion

- When introducing restriction enzyme sites into primers for subsequent digestion and cloning of a PCR product, refer to [www.thermoscientific.com/fd](http://www.thermoscientific.com/fd), Reaction Conditions Guide, to define the number of extra bases required for efficient cleavage..
- Use Thermo Scientific GeneJET PCR Purification Kit, #K0701 to purify PCR product prior digestion in following cases:
  - When PCR additives such as DMSO or glycerol were used, as they may affect the cleavage efficiency or cause star activity.
  - When PCR Product will be used for cloning. Active thermophilic DNA polymerase still present in PCR mixture may alter the ends of the cleaved DNA and reduce the ligation efficiency.

## Activity of DNA Modifying Enzymes in FastDigest and FastDigest Green Buffers, %

Thermo Scientific FastAP Thermosensitive Alkaline Phosphatase, #EF0651	100
T4 DNA Ligase*, #EL0014	75-100
Klenow Fragment, #EP0051	100
T4 DNA Polymerase, #EP0061	100
T4 Polynucleotide Kinase, #EK0031	100

\* 0.5 mM ATP (#R0441) is required for T4 DNA Ligase activity.

## **PRODUCT USE LIMITATION**

This product is developed, designed and sold exclusively *for research purposes and in vitro use only*. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Please refer to [www.thermoscientific.com/onebio](http://www.thermoscientific.com/onebio) for Material Safety Data Sheet of the product.

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