

Annexin-V-FLUOS Staining Kit

Kit for the detection and quantification of apoptosis and differentiation from necrosis at single cell level, based on Annexin-V-labeling

Cat. No. 11 858 777 001 50 tests Cat. No. 11 988 549 001 250 tests

⚠ Version 9.0 Content version: May 2005 Store at +2 to +8°C

1. Product overview

Kit contents

Vial/	Label	Content/Cat.No.		Use
Cap		11 858 777 001	11 988 549 001	USE
	Annexin-V- Fluorescein	110 µl	500 μl	Ready-to-use
	Propidium iodide	150 μΙ	500 μl	Ready-to-use For the preparation of the Annexin-V-Fluorescein labeling solution
_	Incubation buffer	50 ml HEPES buf- fer	4 × 50 ml HEPES buffer	 Ready-to-use For the dilution of the Annexin- V-Fluorescein solution

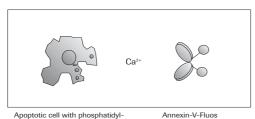
Introduction

In the early stages of apoptosis, changes occur at the cell surface (1, 2, 3). One of these plasma membrane alterations is the translocation of phosphatidylserine (PS) from the inner part of the plasma membrane to the outer layer, by which PS becomes exposed at the external surface of the cell (4). Fadok et al. showed that macrophages specifically recognize PS exposed on the surface of lymphocytes during the development of apoptosis (2). The recognition and phagocytosis of apoptotic cells and bodies protects organisms from the exposure to cellular compounds leading to inflammation, which mostly accompanies necrosis.

Assay principle

The analysis of phosphatidylserine on the outer leaflet of apoptotic cell-membranes is performed by using Annexin-V-Fluorescein and Proidium iodid (PI) for the differentiation from necrotic cells or labeling with a cell surface marker for cell characterization. The procedure involves:

Stage	Description		
1	Washing the cells in PBS.		
2	Incubation of cells with Annexin-V-Fluorescein in a Hepes buffer containing PI or labeling reagent for cell surfaces (e.g., CD-marker).		
3	Analysis of the samples under a fluorescence microscope or on a flow cytometer.		



serine exposed on the outer leaflet of the membrane

Annexin-V-Fluos

Fig. 1: Test principle.

Application

Annexin V is a Ca2+ -dependent phospholipid-binding protein with high affinity for phosphatidylserine (4). This protein can hence be used as a sensitive probe for PS exposure upon the outer leaflet of the cell membrane and is therefore suited to detect apoptotic cells (4, 5, 6, 7) in cell populations but not on tissue sections. Since necrotic cells also expose PS according to the loss of membrane integrity, apoptotic cells have to be differentiated from these necrotic cells. The simultaneous application of a DNA stain which is used for dye exclusion tests allows the discrimination of necrotic cells from the Annexin V positively stained cell cluster. Any other secondary labeling should be possible, *e.g.*, membrane surface staining with a phycoerythrin or TRITC-labeled monoclonal antibody for further cellular characterization (8).

Sample material

- Freshly isolated cells

Number of tests

50 tests (Cat. No. 11 858 777 001) For 250 tests (Cat. No. 11 988 549 001)

Preparation

Recombinant Annexin- V is produced in E. coli (strain NB42). The GST-tagged protein is purified by standard purification protocols.

Fluorescence characteristics

Annexin-V-Fluorescein and propidium iodide show the following fluorescence characteristics:

	Fluorescein	Propidium Iodide
Excitation	488 nm	488-540 nm
Emission	518 nm	617 nm

Specificity

Annexin-V-Fluos binds in a Ca2+-dependent manner to negatively charged phospholipid surfaces and shows high specificity to phosphatidylserine. Therefore, it stains apoptotic as well as necrotic cells. Propidium iodide stains DNA of leaky necrotic cells only.

Storage/Stability

Stable at +2 to +8°C until the expiration date printed on the label.

2. Procedures and required material

Additional solutions required PBS*

Preparation of Annexin-V-FLUOS labeling solution

Predilute 20 µl Annexin-V-Fluos labeling reagent (vial 1) in 1 ml Incubation buffer (bottle 3) and add 20 µl Propidium iodide solution (vial 2). **Note**: 1 ml is enough for 10 samples.

Staining of cell suspensions

In the following table please find the staining procedure for cell suspensions.

Step	Action			
1	Wash 10 6 cells with PBS and centrifuge cells at 200 \times g for 5 min			
	Resuspend the cell pellet in 100 µl of Annexin-V-FLUOS labeling solution. Incubate 10–15 min at 15–25°C.			
3	Analyze by fluorescence microscopy or on a flow cytometer (see 3. Analysis).			

^{*} available from Roche Applied Science

Staining of adherent cells

In the following table please find the staining procedure for adherent cells

Step	Action			
1	Remove chambers and silicon borders of cells grown on chamberslides.			
2	Remove medium and cover slides with Annexin-V-FLUOS labeling solution (100 μl/chamber).			
3	Put coverslips on slides and incubate for 10–15 min at 15–25°C.			
4	Analyze by fluorescence microscopy or on a flow cytometer (see 3. Analysis). Note: We do not recommend to analyze adherent cells by flow cytometry, because trypsinization or scraping for monodispertion of the cells results in false positive staining and analysis of non-dispersed cell clusters.			

3. Analysis

Fluorescencemicroscopy

For evaluation by fluorescence microscopy use an excitation wavelength in the range of 450-500 nm (e.g., 488 nm) and detection in the range of 515-565 nm (green).

Flowcytometry

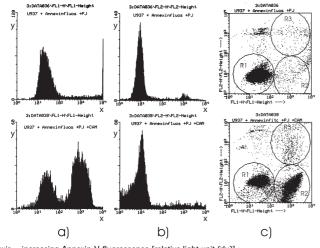
Add 0.5 ml Incubation buffer (bottle 3) per 106 cells and analyze on a flow cytometer using 488 nm excitation and a 515 nm bandpass filter for fluorescein detection and a filter >600 nm for PI detection. Electronic compensation of the instrument is required to exclude overlapping of the two emission spectra. Typical histograms of apoptotic versus non-apoptotic and necrotic cells are shown in figure 2.

Figure 2

FACS analysis of apoptotic U937 cells after staining with Annexin-V-FLUOS and propidium iodide. Cells were then stained with the components of the Annexin-V-FLUOS Staining Kit and analyzed. Cultivation for 4 h in the presence (lower row) or absence (upper row) of 4 µg/ml camptothecin: a) single parameter Annexin-V-FLUOS

b) single parameter propidium iodide and

c) dual parameter (FL1 = Annexin-V-FLUOS; FL2 = propidium iodide); Cluster R1 = living cells, R2 = apoptotic cells and R3 = necrotic cells.



x-axis = increasing Annexin-V-fluorescence [relative light unit (rlu)] v-axis = increasing Propidium Iodide (rlu)

Result: Flow cytometric analysis clearly differentiates normal (living) cells with low Annexin and low PI staining, apoptotic cells with high Annexin and low PI staining, and necrotic cells with high Annexin and high PI staining.

Changes to previous version

Editorial changes

4. References

- Andree, H.A.M. et al. (1990), J. Biol. Chem. 265, 4923.
- Fadok, V. et al. (1992) J. Immunology 148, 2207
- Creutz, C.E. (1992) Science 258, 924
- Vermes, I. et al. (1995) *J. Immunol. Methods* **184**, 39. Koopman, G. et al (1994) *Blood* **84**, 1415.
- Homburg, C.H.E. et al (1995) Blood 85, 532
- Verhoven, B. et al (1995) J. Exp. Med. 182, 1597
- van Engeland, M. et al. (1996) Cytometrie 24, 131.

5. Related products

This table only shows a selection of the most important products related to the product described in this pack insert.

For further information please access our Apoptosis special interest site:

http://www.roche-applied-science.com/apoptosis

Apoptosis- specific physiologi- cal change		Pack size	Cat. No.		
DNA frag-	Gel Electrophoresis				
mentation	Apoptotic DNA-Ladder Kit	20 tests	11 835 246 001		
	In situ assay				
	<i>In Situ</i> Cell Death Detection Kit, TMR red	1 kit (50 tests)	12 156 792 910		
	In Situ Cell Death Detection Kit, Fluorescein	1 kit (50 tests)	11 684 795 910		
	In Situ Cell Death Detection Kit, AP	1 kit (50 tests)	11 684 809 910		
	<i>In Situ</i> Cell Death Detection Kit, POD	1 kit (50 tests)	11 684 817 910		
	Single reagents for TUNEL and		ents		
	TUNEL AP	70 tests (3.5 ml)	11 772 457 001		
	TUNEL POD	70 tests (3.5 ml)	11 772 465 001		
	TUNEL Enzyme	2 × 50 μl (20 tests)	11 767 305 001		
	TUNEL Label Mix	3 × 550 μl (30 tests)	11 767 291 910		
	ELISA				
	Cell Death Detection ELISA	1 kit	11 544 675 001		
	Cell Death Detection ELISAPLUS	1 kit (96 tests)	11 774 425 001		
	Cell Death Detection ELISAPLUS, 10×	1 kit	11 920 685 001		
	Cellular DNA Fragmentation ELISA	1 kit (500 tests)	11 585 045 001		
Cell .	Microscopy or FACS				
membrane	Annexin-V-Alexa 568	250 tests	03 703 126 001		
alterations	Annexin-V-Biotin	250 tests	11 828 690 001		
	Annexin-V-FLUOS	250 tests	11 828 681 001		
	Annexin V FLUOS Staining Kit	50 tests 250 tests	11 858 777 001 11 988 549 001		
Enzymatic	Western Blot				
activity	Anti-Poly (ADP-Ribose) Poly- merase	100 μl	11 835 238 001		
	FIENA				
	Caspase 3 Activity Assay	1 kit	12 012 952 001		
	Fluorimetric microplate Assay				
	Homogeneous Caspase Assay, fluorometric	100 tests 1000 tests	03 005 372 001 12 236 869 001		
	In situ Assay				
	M30 CytoDEATH (formalin grade)	50 tests 250 tests	12 140 322 001 12 140 349 001		
	M30 CytoDEATH, Fluorescein	250 tests	12 156 857 001		
Expression	Apoptosis Induction				
of apopto-	Anti-Fas (CD95/APO-1)	1000 tests	11 922 432 001		
sis-related	ELISA				
proteins	p53 pan ELISA	1 kit	11 828 789 001		

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