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Q5® Site-Directed Mutagenesis (E0554) V.1 [↗](#)

1 Works for me [dx.doi.org/10.17504/protocols.io.cjpumm](https://doi.org/10.17504/protocols.io.cjpumm)

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ABSTRACT

This is the protocol for the Q5® Site-Directed Mutagenesis Kit (E0554)

EXTERNAL LINK

<https://www.neb.com/protocols/2013/01/26/q5-site-directed-mutagenesis-kit-protocol-e0554>

GUIDELINES

DESCRIPTION

The Q5® Site-Directed Mutagenesis Kit enables rapid, site-specific mutagenesis of double-stranded plasmid DNA in less than 2 hours (Figure 1). The kit utilizes the robust Q5 Hot Start High-Fidelity DNA Polymerase along with custom mutagenic primers to create insertions, deletions and substitutions in a wide variety of plasmids. After PCR, the amplified material is added directly to a unique Kinase-Ligase-DpnI (KLD) enzyme mix for rapid (5 minutes), room temperature circularization and template removal (Figure 2). Transformation into high-efficiency NEB 5-alpha Competent E. coli, provided with the kit, ensures robust results with plasmids up to at least 20 kb in length.

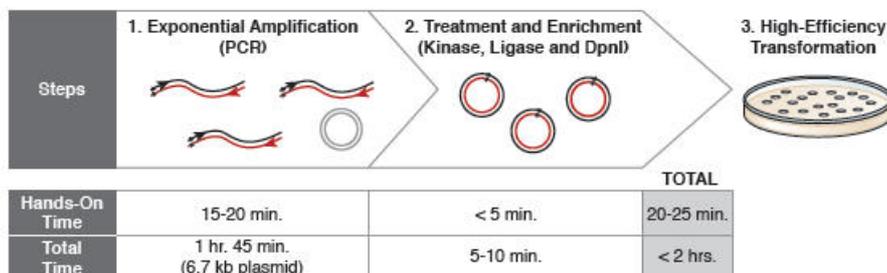


Figure 1: Site-specific mutagenesis proceeds in less than 2 hours.

The use of a master mix, a unique multi-enzyme KLD enzyme mix, and a fast polymerase ensures that, for most plasmids, the mutagenesis reaction is complete in less than two hours.

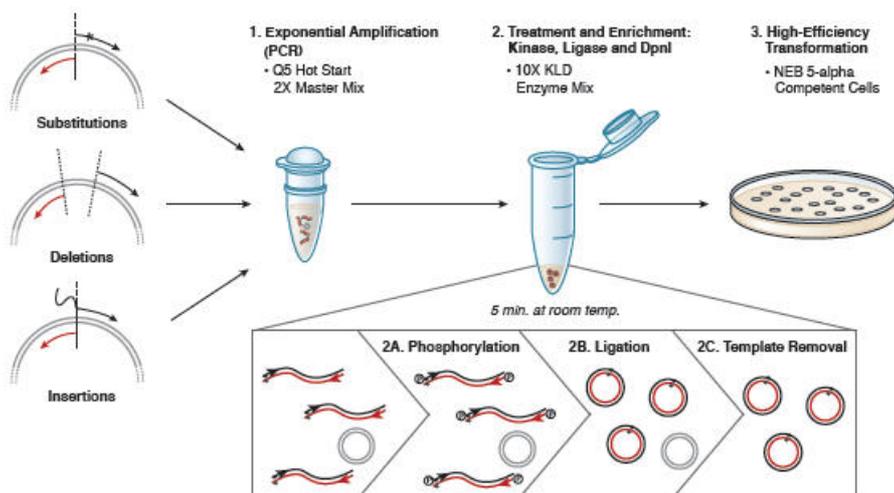


Figure 2: Q5 Site-Directed Mutagenesis Kit Overview.

This kit is designed for rapid and efficient incorporation of insertions, deletions and substitutions into doublestranded plasmid DNA. The first step is an exponential amplification using standard primers and a master mix formulation of Q5 Hot Start High-Fidelity DNA Polymerase. The second step involves incubation with a unique enzyme mix containing a kinase, a ligase and DpnI. Together, these enzymes allow for rapid circularization of the PCR product and removal of the template DNA. The last step is a high-efficiency transformation into chemically competent cells (provided).

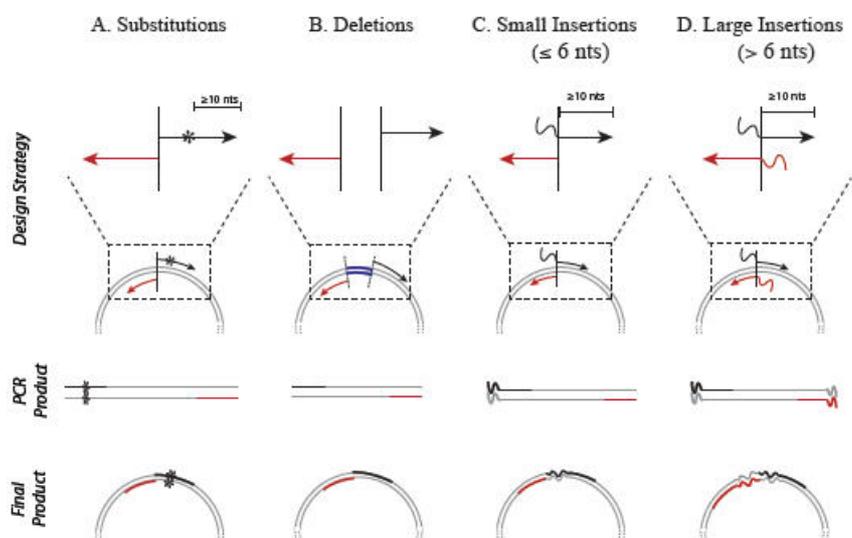


Figure 3: Primer Design for the Q5 Site-Directed Mutagenesis Kit

Substitutions, deletions and insertions are incorporated into plasmid DNA through the use of specifically designed forward (black) and reverse (red) primers. **Unlike kits that rely on linear amplification, primers designed for the Q5 Site-Directed Mutagenesis Kit should not overlap to ensure that the benefits of exponential amplification are realized.** A) Substitutions are created by incorporating the desired nucleotide change(s) (denoted by *) in the center of the forward primer, including at least 10 complementary nucleotides on the 3' side of the mutation(s). The reverse primer is designed so that the 5' ends of the two primers anneal back-to-back. B) Deletions are engineered by designing standard, non-mutagenic forward and reverse primers that flank the region to be deleted. C) Insertions less than or equal to 6 nucleotides are incorporated into the 5' end of the forward primer while the reverse primer anneals back-to-back with the 5' end of the complementary region of the forward primer. D) Larger insertions can be created by incorporating half of the desired insertion into the 5' ends of both primers. The maximum size of the insertion is largely dictated by oligonucleotide synthesis limitations.

TROUBLESHOOTING

No/Low Colonies

- Ensure that your primers are designed properly. To take advantage of the exponential nature of the amplification reaction, the 5' ends of the two primers should align back-to-back unless deletions are being made (see Figure 3). For best results, primers should be designed and annealing temperatures calculated using [NEBaseChanger™](#), the NEB online primer design software.
- Ensure there is a clean PCR product by visualizing 2–5 µl of the reaction on an agarose gel. Follow the suggestions below for low or impure PCR products.
- Only use 1 µl of PCR product in the KLD reaction. Carrying too much PCR product forward can decrease transformation efficiency. If the PCR yield is low, more product can be added to the KLD reaction, however a buffer exchange step, such as PCR purification, must be included prior to transformation.
- Only use 5 µl of the KLD reaction in the transformation. If more KLD reaction is added, a buffer exchange step, such as PCR purification, should be included prior to transformation.
- Ensure that the selectable marker in the plasmid matches the selection agent used in the plates
- Ensure the NEB 5-alpha Competent E. coli cells have been stored at -80° C.
- Check that the transformation efficiency of the competent cells is ~1 x 10⁹ colony forming units (cfu) per µg. To calculate transformation efficiency, transform 2 µl of the provided control pUC19 DNA (100 pg) into 50 µl of cells. Follow the transformation protocol on page 8. Prior to plating, dilute 10 µl of cells up to 1 ml in SOC. Plate 100 µl of this dilution. In this case, 150 colonies will yield a transformation efficiency of 1.5 x 10⁹ cfu/µg
- (µg DNA=0.0001, dilution=10/1000 x 100/1000).

No/Low PCR Product

- Ensure that the optimal annealing temperature (Ta) is used.
- High-Fidelity polymerases benefit from a Tm+3 annealing temp. Use [NEBaseChanger™](#), the NEB online primer design software, to calculate Ta. Alternatively, the optimal annealing temperature could be determined using a gradient PCR followed by agarose gel analysis.
- Ensure that the elongation time is adequate for the plasmid length. We recommend 10–20 seconds per kb of plasmid.
- Ensure that the final concentration of each primer is 0.5 µM.
- Purify the primers with polyacrylamide gel electrophoresis (PAGE).

Resulting Plasmids Do Not Contain the Desired Mutation

- Ensure proper design of the mutagenic primers.
- Optimize the PCR conditions (see above).
- Use 1–25 ng of template in the PCR step. A small increase in the number of clones with no/incorrect mutation incorporated can occur if less than 1 ng or more than 25 ng of template is used.

NOTES

Storage Note:

The Q5 Site-Directed Mutagenesis Kit is stable at -80°C for one year. For convenience, the Q5 Hot Start High-Fidelity 2X Master Mix, KLD Enzyme Mix, KLD Reaction Buffer, Control Primers and Template DNA are packaged together in a separate box that can be removed and stored at -20°C for two years with no loss of activity. The SOC can be removed and stored at room temperature. It is important to store the NEB 5-alpha Competent E. coli at -80°C, and avoid repeated freeze-thaw cycles.

REFERENCES

1. Kalnins et al., (1983). The EMBO Journal. 2, 593-597.
2. Dickinson DJ, Ward JD, Reiner DJ, Goldstein B. (2013). Engineering the Caenorhabditis elegans genome using Cas9-triggered homologous recombination.. Nat Methods. Sep 1, PubMedID: 23995389

MATERIALS

NAME ▾	CATALOG # ▾	VENDOR ▾
Q5 Site-Directed Mutagenesis Kit - 10 rxns	E0554S	New England Biolabs

STEPS MATERIALS

NAME ▾	CATALOG # ▾	VENDOR ▾
SOC Outgrowth Medium - 100 ml	B9020S	New England Biolabs

Exponential Amplification (PCR)

1 Assemble the following reagents in a thin-walled PCR tube.

	25 μ l RXN	FINAL CONC.
Q5 Hot Start High-Fidelity 2X Master Mix	12.5 μ l	1X
10 μ M Forward Primer	1.25 μ l	0.5 μ M
10 μ M Reverse Primer	1.25 μ l	0.5 μ M
Template DNA (1–25 ng/ μ l)	1 μ l	1–25 ng
Nuclease-free water	9.0 μ l	




E0552 Q5 PCR Mixture
by New England Biolabs,
New England Biolabs

PREVIEW

RUN

1.1 Q5 Hot Start High-Fidelity 2X Master Mix **12.5 μ l**

1.2 10 μ M Forward Primer **1.25 μ l**

1.3 10 μ M Reverse Primer **1.25 μ l**

1.4 Template DNA (1–25 ng/ μ l) **1 μ l**

1.5 Nuclease-free water **9 μ l**

2 Mix reagents completely.

3 Transfer to a thermocycler and perform the following cycling conditions:

Thermocycling Conditions for a Routine PCR:

STEP	TEMP	TIME
Initial Denaturation	98°C	30 seconds
25 Cycles	98°C	10 seconds
50–72°C*	10–30 seconds	2 minutes
72°C	20–30 seconds/kb	
Final Extension	72°C	
Hold	4–10°C	

COMMENTS

For a Q5-optimized annealing temperature of mutagenic primers, please use [NEBaseChanger™](#), the online NEB primer design software. For pre-designed, back-to-back primer sets, a $T_a = T_m + 3$ rule can be applied, but optimization may be necessary.

Kinase, Ligase & DpnI (KLD) Treatment

4 Assemble the following reagents:



E0552 KLD Mixture
by New England Biolabs,
New England Biolabs

[PREVIEW](#) [RUN](#)

4.1 PCR Product **1ul**

4.2 2X KLD Reaction Buffer **5ul**

4.3 10X KLD Enzyme Mix **1ul**

4.4 Nuclease-free Water **3ul**

5 Mix well by pipetting up and down.

6 Incubate at room temperature for 5 minutes.

 00:05:00

Transformation

7 Thaw a tube of NEB 5-alpha Competent *E. coli* cells on ice.

8 Add **5 µl** of the KLD mix from the "KLD Section" above to the tube of thawed cells.

9 Carefully flick the tube 4-5 times to mix. **Do not vortex.**

10 Place the mixture on ice for 30 minutes.

 00:30:00

11 Heat shock at 42°C for 30 seconds.

 00:00:30

12 Place on ice for 5 minutes.

 00:05:00

13 Pipette 950 μ l of room temperature SOC into the mixture.

 950 μ l

 **SOC Outgrowth Medium - 100 ml**
by [New England Biolabs](#)
Catalog #: [B9020S](#)

14 Incubate at 37°C for 60 minutes with shaking (250 rpm).

 01:00:00

15 Mix the cells thoroughly by flicking the tube and inverting.

COMMENTS

[New England Biolabs](#) Jan 26, 2015 10:59 PM
[New England Biolabs](#) edited on Jan 26, 2015 10:59 PM

It may be necessary (particularly for simple substitution and deletion experiments) to make a 10- to 40-fold dilution of the transformation mix in SOC prior to plating, to avoid a lawn of colonies

16 Spread 50-100 μ l onto a selection plate.

17 Incubate overnight at 37°C

 15:00:00



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