



Product Manual

GFP CERTIFIED® **Apoptosis/Necrosis Detection Kit**

Instruction Manual

Catalog Number: ENZ-51002

ENZ-51002-25

25 assays

ENZ-51002-100

100 assays



Product Manual

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Please read entire booklet before proceeding with the assay.



Carefully note the handling and storage conditions of each kit component.



Please contact Enzo Life Sciences Technical Support if necessary.

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INTRODUCTION

The transition from apoptosis to necrosis is a loosely defined continuum that necessitates recognition of the various stages of the process. The display of phosphatidylserine (PS) on the extracellular face of the plasma membrane remains a unifying hallmark of early apoptosis. Phospholipid-binding proteins such as Annexin V bind with a high affinity to PS, in the presence of Ca^{2+} . Given that Annexin V is not cell permeable, the binding of externalized PS is selective for early apoptotic cells. Similarly, loss of plasma membrane integrity, as demonstrated by the ability of a membrane-impermeable DNA intercalating dye to label the nucleus, represents a straightforward approach to demonstrate late stage apoptosis and necrosis.

Considering that Annexin V is commonly conjugated to fluorescein (a.k.a. FITC), apoptosis detection in green fluorescent protein (GFP)-expressing cells can be problematic. The spectral overlap between the emission profiles of these two fluorophores makes differentiation between signals difficult or impossible, thus compromising data quality overall. While useful for analysis of apoptosis and necrosis in cells that do not express any fluorescent proteins, the GFP-CERTIFIED[®] Apoptosis/Necrosis Detection Kit was specifically designed for use with GFP-expressing cell lines and cells expressing blue or cyan fluorescent proteins (BFPs, CFPs). Additionally, the kit is suitable for use with live or post-fixed cells in conjunction with probes, such as labeled antibodies, or other fluorescent conjugates displaying similar spectral properties as fluorescein or coumarin.

Each GFP CERTIFIED[®] Apoptosis/Necrosis Detection Kit includes all the necessary reagents for determination of early and late stages of apoptosis as well as necrosis. An Annexin V-EnzoGold (enhanced Cyanine 3) conjugate enables detection of apoptosis distinct from fluorescein or GFP. The Necrosis Detection Reagent similar to the red-emitting dye 7-AAD, facilitates late apoptosis and necrosis detection. This kit also includes an Apoptosis Inducer (Staurosporine) for use as a positive control. Detection reagents can be used separately or in combination for either single or multiplexed applications respectively. This kit also provides quick and accurate results via flow cytometry and fluorescence/confocal microscopy.

SAFETY WARNINGS & PRECAUTIONS



Avoid freeze /
thaw cycles



Handle with
care

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

- This product is for research use only and is not intended for diagnostic purposes.
- Some components of this kit contain hazardous substances. Reagents can be harmful if ingested or absorbed through the skin and may cause irritation to the eyes. Reagents should be treated as possible mutagens and should be handled with care and disposed of properly.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas. All blood components and biological materials should be treated as potentially hazardous and handled as such. They should be disposed of in accordance with established safety procedures.
- To avoid photobleaching, perform all manipulations in low light environments, in amber microcentrifuge tubes or protected from light by other means.

MATERIALS SUPPLIED

All reagents are shipped on wet ice. Upon receipt, store the Annexin V-EnzoGold and Binding Buffer at 4°C, protected from light. Store all other reagents at -20°C, protected from light.

Reagent	ENZ-51002-100	ENZ-51002-25
Apoptosis Detection Reagent (Annexin V-EnzoGold)	550 µL	140 µL
Necrosis Detection Reagent	600 µL	150 µL
Apoptosis Inducer (Staurosporine)	50 nmoles	50 nmoles
Binding Buffer (10X)	6 mL	1.5 mL

ADDITIONAL MATERIALS REQUIRED

- Standard fluorescence microscope or a flow cytometer that is equipped with a 488 nm laser for fluorescent dye excitation
- Phosphate buffered saline (PBS)
- Calibrated, adjustable precision pipetters, preferably with disposable plastic tips
- 5mL tubes appropriate for holding cells during induction of apoptosis and necrosis
- Adjustable speed centrifuge with swinging buckets
- Glass microscope slides (for microscope analysis only)
- Glass cover slips (for microscope analysis only)
- Deionized water
- Anhydrous DMSO

METHODS AND PROCEDURES

A. REAGENT PREPARATION

1. Positive Control

The Apoptosis Inducer (Staurosporine) is supplied as a lyophilized powder (50nmol) and should be reconstituted in 50 μ L DMSO for a 1mM stock solution. It is recommended that induction occur at the effective concentration (EC50) and that the final percent DMSO in the assay not exceed 0.2%.

2. Cell Preparation

Cells should be maintained via standard tissue culture practices. Positive Control cells should be pretreated with the Apoptosis Inducer for 1-16 hours. Induction of apoptosis and necrosis is time and concentration dependent and may also vary significantly depending among cell types and cell lines. Post treatment cells should be harvested and diluted to a working concentration of 2.0×10^5 /mL in media. Negative control cells should be treated with vehicle (DMSO, media or other solvent used to reconstitute or dilute an inducer or inhibitor) for an equal length of time under similar conditions.

3. 1X Binding Buffer

Generate 500 μ L of 1X Binding Buffer per sample by diluting 50 μ L of 10X Binding Buffer into 450 μ L of deionized water. Batch preparation is recommended for large numbers of samples.

B. DUAL DETECTION REAGENT PREPARATION

Table 1. Dual Detection Reagent

Prepare the Dual Detection Reagent according to Table 1. The volumes recommended in the table are sufficient for one assay and must be scaled accordingly.

Reagent	Amount
1X Binding Buffer	500 μ L
Apoptosis Detection Reagent (Annexin V-EnzoGold)	5 μ L
Necrosis Detection Reagent	5 μ L
Total Volume	510 μL

C. FLOW CYTOMETRY PROTOCOL (Non-Adherent Cells)

1. The recommended sample size should be between 2×10^5 cells/sample and 1×10^6 cells/sample during staining and analysis.
2. Treat cells with compound of interest and Negative Control cells with vehicle.
3. Centrifuge cells for 5 minutes at $400 \times g$ at room temperature (RT).
4. Carefully remove supernatant by aspiration and gently resuspend and wash the pellet in 5.0 mL cold PBS.
5. Centrifuge cells for 5 minutes at $400 \times g$ at room temperature (RT).
6. Carefully remove supernatant and gently resuspend the pellet in 500 μ L Dual Detection Reagent. (see **Table 1** in section B.)
7. Protect samples from light and incubate for 15 minutes at room temperature.
8. Samples should be analyzed via flow cytometry using a 488nm laser with the FL 1 channel for GFP or FITC detection, the FL 2 channel for the Apoptosis Detection Reagent and the FL 3 channel for the Necrosis Detection Reagent. It is recommended that compensation corrections be performed using single stained induced cells to avoid any overlap between the Apoptosis Detection Reagent and Necrosis Detection Reagent fluorescent signals.
9. Positive Control Samples: It is recommended that positive control samples be pretreated with the Apoptosis Inducer (Staurosporine) at a final concentration of 2μ M for 4 hours. Apoptosis may vary significantly and is dependent upon the length of induction, inducer concentration, cell types, cell lines, inducer concentration and exposure time. Follow steps 2-8 post treatment.

Suggested Compensation Controls for Flow Cytometry:

- a. Unstained cells
- b. Cells stained with Apoptosis Detection Reagent (without Necrosis Detection Reagent)
- c. Cells stained with Necrosis Detection Reagent (without Apoptosis Detection Reagent)

Note: Samples and controls should be kept on ice before the assay is run and analyzed via flow cytometry within one hour of staining.

D. FLOW CYTOMETRY PROTOCOL (Adherent Cells)

1. The recommended sample size should be between 2×10^5 cells/sample and 1×10^6 cells/sample during staining and analysis.
2. Treat cells with compound of interest and Negative Control cells with vehicle.
3. Detach cells via trypsinization or other cell dissociation method. Minimize cellular damage by optimizing the concentration and time cells are exposed to Trypsin.
4. Centrifuge cells for 5 minutes at $400 \times g$ at room temperature (RT).
5. Carefully remove supernatant by aspiration and gently resuspend and wash the pellet in 5.0 mL cold PBS.
6. Centrifuge cells for 5 minutes at $400 \times g$ at room temperature (RT).
7. Carefully remove supernatant and gently resuspend the pellet in 500 μ L Dual Detection Reagent (see **Table 1** in section B.).
8. Protect samples from light and incubate for 15 minutes at room temperature.
9. Samples should be analyzed via flow cytometry using a 488nm laser with the FL 1 channel for GFP or FITC detection, the FL 2 channel for the Apoptosis Detection Reagent and the FL 3 channel for the Necrosis Detection Reagent. It is recommended that compensation corrections be performed using single stained induced cells to avoid any overlap between the Apoptosis Detection Reagent and Necrosis Detection Reagent fluorescent signals.
10. Positive Control Samples: It is recommended that positive control samples be pretreated with the Apoptosis Inducer at a final concentration of 2 μ M for 4 hours. Apoptosis may vary significantly and is dependent upon the length of induction, inducer concentration, cell types, cell lines, inducer concentration and exposure time. Follow steps 2-9 post treatment.
11. Suggested Compensation Controls for Flow Cytometry:
12. Unstained cells
13. Cells stained with Apoptosis Detection Reagent (without Necrosis Detection Reagent)

14. Cells Stained with Necrosis Detection Reagent (without Apoptosis Detection Reagent)

Note: Samples and controls should be kept on ice before the assay is run and analyzed via flow cytometry within one hour of staining.

Suggested Compensation Controls for Flow Cytometry:

- a. Unstained cells
- b. Cells stained with Apoptosis Detection Reagent (without Necrosis Detection Reagent)
- c. Cells Stained with Necrosis Detection Reagent (without Apoptosis Detection Reagent)

Note: Samples and controls should be kept on ice before the assay is run and analyzed via flow cytometry within one hour of staining.

E. WIDE FIELD FLUORESCENCE/CONFOCAL MICROSCOPY

1. Grow cells directly onto glass slides or polystyrene tissue culture plates until ~80% confluent.
2. Treat cells with compound of interest and Negative Control cells with vehicle.
3. Carefully wash cells twice with 1x PBS in a volume sufficient for covering the cell monolayer.
4. Carefully remove supernatant and dispense the Dual Detection Reagent in a volume sufficient for covering the cell monolayer (as per section B)
5. Protect samples from light and incubate for 15 minutes at room temperature.
6. Flick the staining solution onto a paper towel and if necessary add a few drops of Binding Buffer to prevent the cells from drying out.
7. Cover cells and observe under a fluorescence/confocal microscope with a dual filter set for Cyanine-3 (Ex/Em: 550/570 nm) and 7-AAD (Ex/Em: 546/647 nm) and a GFP/FITC (Ex/Em: 488/514 nm) filter should be used if either is imaged simultaneously.
8. Positive Control Samples: It is recommended that Positive Control samples be pretreated with the Apoptosis Inducer at a final concentration of 2 μ M for 4 hours and may vary significantly among cell types and cell lines. Follow steps 3-7 post treatment.

APPENDICES

A. FILTER SELECTION

The selection of optimal filter sets for a fluorescence microscopy application requires matching the optical filter specifications to the spectral characteristics of the dyes employed in the analysis. Please consult your microscope or filter set manufacturer for assistance in selecting optimal filter sets for your microscope.

B. CELL FIXATION

- **IMPORTANT!** The fixation process can disrupt cellular membranes and should be performed following incubation with the Detection Reagent.
- 2% formaldehyde is the recommended fixative.
- Standard immunofluorescence staining protocols using a fluorescein- or coumarin-based antibody conjugates, or equivalent should be administered post-fixation according to manufacturer instructions.

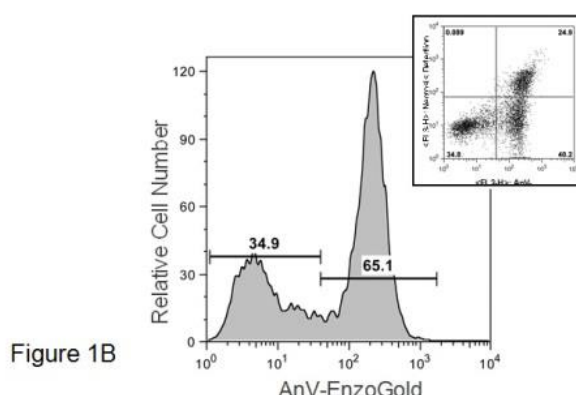
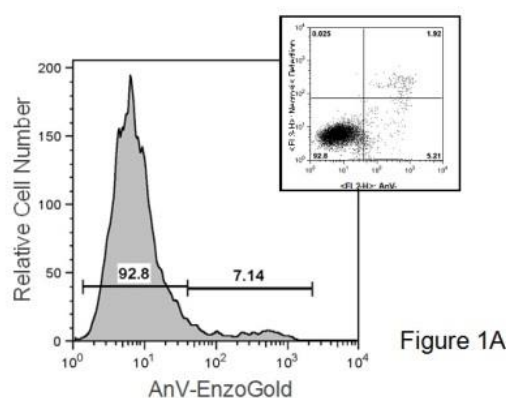
C. RESULTS

Apoptosis is characterized by a diverse repertoire of biochemical and cellular signals that alter cellular morphology. Some of the downstream effects observed via fluorescence microscopy include cell shrinkage, loss of membrane symmetry, membrane blebbing and cytoplasmic condensation. Changes at the nuclear level are also evident, including the condensation and fragmentation of nuclear and chromosomal DNA. By contrast, necrosis results from acute cellular damage or damage to an essential cellular system and is morphologically defined by early plasma membrane rupture and dilatation of organelles.

Cells treated with Staurosporine or other inducers demonstrate four possible cell populations via fluorescence microscopy. Viable cells are negative for both the Apoptosis Detection Reagent (Cyanine-3) and Necrosis Detection Reagent (far-red). Apoptotic cells with are positive for the Apoptosis Detection Reagent while cells positive for both the Apoptosis Detection Reagent and Necrosis Detection Reagent demonstrate late-stage apoptosis. Finally, necrotic cells are positive for the Necrotic Detection Reagent and Apoptosis Detection reagent negative.

Typical results of flow cytometry-based analysis of cell populations using the Enzo GFP CERTIFIED® Apoptosis/Necrosis detection kit. Control populations of Jurkat (T-Cell leukemia) cells stained but not induced are negative for either detection reagent with the exception of a small percentage (up to 20%) as expected in routine cultures of untreated cells (Figure 1A). Cells treated with 2 μ M Staurosporine for 4 hours show cells positive for the Apoptosis Detection Reagent (apoptosis) and cells positive for both the Apoptosis Detection Reagent and Necrosis Detection Reagent (late-stage apoptosis).

(Figure 1B).



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3. van Engeland M, Ramaekers FC, Schutte B, Reutelingsperger CP. A novel assay to measure loss of plasma membrane asymmetry during apoptosis of adherent cells in culture. *Cytometry.* 1996;24(2):131-9.
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an apoptosis detection system based on phosphatidylserine exposure. Cytometry. 1998;31(1):1-9. Review.

TROUBLESHOOTING GUIDE

Problem	Potential Cause	Suggestions
Little or no apoptosis observed or detected	No or few apoptotic cells present in sample	Use the Apoptosis Inducer, provided in the kit, as a positive control to induce apoptosis and optimize the Inducer concentration and time of treatment.
	Incorrect buffer used during sample processing	Calcium and magnesium ions are required for binding of the Apoptosis Detection Reagent to PS. Binding is reversible, so divalent cations must be present throughout the entire assay. Use the Binding Buffer provided in the kit.
		Completely remove wash buffer by aspiration as to not dilute the binding buffer or inducer.
Significant apoptosis detected in negative control cells.	Assay is inappropriate for the cells or cell line used.	Some cells (<i>i.e.</i> , megakaryocytes, platelets, some myeloid lineage cells) are known to display large amounts of PS on their cell surfaces. Peripheral Blood Mononuclear Cells (PBMCs) coated with platelets, therefore, may be positive for the Apoptosis Detection Reagent. Use a different method to monitor apoptosis.
	Cells were damaged or permeabilized during the assay	Permeable cells (advanced apoptotic or necrotic), yield an increase in apoptosis detection via cytoplasmic leaflet of the plasma membrane. Trypsinization or scraping or disaggregation of adherent cells may result in false positives due to cell damage, arising from disruption of the PS-containing plasma membrane. Use gentler cell preparation methods and optimize the Inducer concentration and time of treatment.
	Assay measurement or analysis was delayed.	Analysis must be performed quickly, following dye labeling in order to avoid post-label apoptosis. Stained samples should be kept on ice until an assay is run.

Problem	Potential Cause	Suggestions
Small apoptotic population detected and high necrotic population detected	Cells are no longer apoptotic and have progressed towards necrotic	High doses of an agent can often lead to necrosis, while lower doses of the same agent may lead to apoptosis. Often times, apoptosis and necrosis simply represent two extremes of biochemically overlapping cell death pathways. Lower the dose of the test agent.
High variability of results observed between and among samples on the same day or in different experiments from day to day	Inconsistencies in cell treatment, labeling and FCM data analysis were introduced.	All reagents should be prepared in batches. An untreated negative control and if possible an independent positive control should be included. Proper controls should be included in each experiment and gating of samples should be performed uniformly especially the gating out of debris.
Spectral emission overlap or signal bleed among different channels or emission filters is observed	No or incorrect compensation was performed for flow cytometry	Use correct compensation controls for flow cytometry and set up spectral compensation according to the manufacturer's instructions.
	Inappropriate filter set used with fluorescence microscopy	Use correct filters for fluorophores being measured. Refer to the spectral chart in this manual.
		Decrease band width of the emission filters to prevent spectral overlap.



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NOTES



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GLOBAL HEADQUARTERS

Enzo Life Sciences Inc.
10 Executive Boulevard
Farmingdale, NY 11735
Toll-Free: 1.800.942.0430
Phone: 631.694.7070
Fax: 631.694.7501
info-usa@enzolifesciences.com

EUROPE/ASIA

Enzo Life Sciences (ELS) AG
Industriestrasse 17
CH-4415 Lausen
Switzerland
Phone: +41/0 61 926 89 89
Fax: +41/0 61 926 89 79
info-ch@enzolifesciences.com

For local distributors and detailed product information visit us online:
www.enzolifesciences.com