

# XcelGen | Plant RNA Mini Kit User Guide

Cat No: XG6611-01 XcelGen Ver.: 0.2/13 Revised Protocol



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#### Introduction

The Plant RNA Mini Kit provides an easy and fast method for isolating RNA from Plant tissue within 30 min. Only trace genomic DNA exists in the purified RNA, which can be eliminated by DNase I treatment when it is necessary.

#### **Overview**

If using the Plant RNA Mini Kit for the first time, please read this booklet to become familiar with the procedures. Samples are homogenized and lysed in a high salt buffer. The RNA is bound to the column while DNA, proteins and other impurities are removed by wash buffer.

#### **Storage and Stability**

All components of the Plant RNA Kit are stable for at least 12 months when stored at  $22^{\circ}$ C- $25^{\circ}$ C. During shipment, or storage in cool ambient conditions, precipitates may form in Buffer RLY. It is possible to dissolve such deposits by warming the solution at  $50^{\circ}$ C, though we have found that they do not interfere with overall performance.



#### **Kit Contents**

Product	XG6611-00	XG6611-01
Preps	4	50
Collection Tubes	8	100
Buffer RLY	3 ml	30 ml
Buffer RB	2 ml	20 ml
RNA Wash Buffer*	1.2 ml	24 ml
DEPC-Treated H <sub>2</sub> O	1 ml	5 ml
RNA Columns	4	50
DNA Clearance Column	4	50
1.5ml RNase-free microfuge tube	4	50
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#### **Before Starting**

- Add 4.8 ml (XG6611-00) or 96 ml (XG6611-01) 100% ethanol to each RNA Wash Buffer before use.
- Add 3 ml (XG6611-00) or 30 ml (XG6611-01) 100% ethanol to each Buffer RB before use.

#### Protocol for Extracting Total RNA From Plant Tissue

#### Note: Perform all steps including centrifugation at room temperature

- 1. Weigh **50mg** plant tissue (no more than 100 mg) in a 2 ml tube. Freeze the plant tissue in liquid nitrogen and grind using a rotor starter.
- 2. Transfer **5 volume (500\muI) Buffer RLY/\beta-mercaptoethanol** to the tube containing the plant tissue immediately. Grind using a rotor starter again.

**Note:** Ensure that  $\beta$ -mercaptoethanol has been added before use. **10µl**  $\beta$ -**mercaptoethanol** should be added in **500µl RLY**.

3. Transfer the cleared lysate to a DNA Clearance column pre-inserted in a 2 ml Collection tube. Centrifuge at 13,000 rpm for 2 min. Discard the DNA Clearance column and save the flow-through.



Note: This step is for genomic DNA removal.

- 4. Add **1 volume 100% ethanol** to the lysate (for example: 500µl 100% ethanol for 500µl lysate).
- 5. Transfer the solution into a RNA column and centrifuge at 13,000 rpm for 1 min. Discard the collection tube with the flow-through and put the column back to a new collection tube.
- 6. Add **400µl Buffer RB** to the column and centrifuge at 13,000 rpm for 30s. Discard the flow-through.
- 7. Add **650µl RNA Wash Buffer** to the column and centrifuge at 13,000 rpm for 1 min. Discard the flow-through.

Note: Ensure that ethanol has been added to RNA Wash Buffer before use.

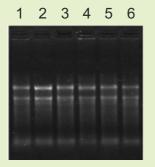
- 8. Add another **650µl RNA Wash Buffer** to the column and centrifuge at 13,000 rpm for 30 s. Discard the flow-through and collection tube, put the column, with the lid open, back to a new collection tube.
- 9. Centrifuge at 13,000 rpm for 5 min. Discard the flow-through.

**Note:** The residual ethanol will be removed more efficiently with the column lid open during centrifugation.

10. Place the column to a RNase-free 1.5 ml tube, add **30-50 µl DEPC-treated ddH<sub>2</sub>O** to the column, incubate at room temperature for 1 min, and centrifuge at 13,000 rpm for 1 min. The RNA is in the flow-through liquid. Store the RNA solution at - 20°C. Reload the eluted RNA solution to the column and centrifuge at 13,000 rpm again.

**Note:** It is highly recommended that RNA quality to be determined before downstream applications. The quality of RNA can be assessed by denatured agarose gel electrophoresis with the ethidium bromide staining. Several sharp bands should appear on the gel including 28S and 18S ribosomal RNA bands as well as certain populations of mRNA and bands. If these bands smear towards lower molecular weight RNAs, then the RNA has undergone major degradation during preparation, handling or storage, RNA molecule less than 200 bases in length do not efficiently bind to the RNA column. An  $A_{260}/A_{280}$  ratio of 1.8-2.0 corresponds to 90-100% pure nucleic acid.





**Fig:** Denaturing Agarose gel analysis of RNA isolated from different plant samples with XcelGen Plant RNA mini Kit.

Lane 1: RNA Isolated from Rice Lane 2: RNA Isolated from Bamboo Lane 3: RNA Isolated from Wheat Lane 4: RNA Isolated from Jasminum Sambac Lane 5: RNA Isolated from Prosopis Lane 6: RNA Isolated from Banana

#### **Troubleshooting Guide**

Problems	Possible Reasons	Suggestion
Low A <sub>260</sub> /A <sub>280</sub> ratio	Protein contamination	Do a Phenol: Chloroform extraction. Loss of total RNA (up to 40%) should be expected.
Low Yield	RNA in sample degraded	Freeze samples immediately in liquid nitrogen and store at -70°C after collect it.
Low Yield	The binding capacity of the membrane in the spin column was exceeded	Use of too much tissue sample exceeding the binding capacity of spin column will cause the decreasing of total RNA yield.
Low Yield	Ethanol not added to Buffer	Add ethanol to the RNA Wash Buffer and Buffer RW before purification.
Genomic DNA contamination	Too much total RNA sample was used in RT-PCR.	Reduce total RNA amount used in RT- PCR to 50-100 ng.
Genomic DNA contamination	The sample may contain too much genomic DNA.	Reduce the amount of starting tissue in the preparation of the homogenate. Most tissues will not show a genomic DNA contamination problem at 30 mg or less per prep.



#### Limited Use and Warranty

This product is intended for in vitro research use only. Not for use in human.

This product is warranted to perform as described in its labeling and in XcelGen's literature when used in accordance with instructions. No other warranties of any kind, express or implied, including, without limitation, implied warranties of merchantability or fitness for a particular purpose, are provided by XcelGen. XcelGen's sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of XcelGen, to replace the products, XcelGen shall have no liability for any direct, indirect, consequential, or incidental damage arising out of the use, the results of use, or the inability to use it product.

For technical support or for more product information, please visit our website at **www.xcelrisgenomics.com** 



### **Product & Services**



Plasmid DNA Isolation Kits • Genomic DNA Extraction Kits • RNA Extraction Kits • Polymerase • DNA Ladders • DNA Markers
 • Premix Tag • dNTP's • RAPD kits • Agarose • Glycerol • Tms NA Stabilizers & RNA Protectant solutions



• 10 nmole • 25 nmole • 50 nmole • 100 nmole • 200 nmole • 1000 nmole



Denovo Genome Sequencing • Whole Genome Resequencing • GBS/RAD Sequencing • Exome Sequencing • Amplicon Sequencing

- Whole Transcriptome Analysis/RNA-Sequencing Small RNA Sequencing Metagenomics Metatranscriptomics
  - ChIP Sequencing 
    Mitochondrial Sequencing 
    Next Generation Genomic Services on Illumina MiSeq
  - Genotyping by Sequencing Tilling/Ecotilling using NGS Genome Database development Services

#### **NGS Bioinformatics**

In silico Primer Design
 Microarray Analysis
 Metagenomics
 Physical, Genetic and QTL mapping
 Assembly and annotation of prokaryotic and eukaryotic genome
 Genome Mapping and SNP discovery
 Transcriptome discovery and analysis
 sRNA analysis and discovery



Plasmid /PCR Sequencing Services • r-E. coli Culture Sequencing Services • Primer Walk Sequencing Services
 Microbial Identification Service • Multilocus Sequence Typing

#### **Customised Services**

SNP Genotyping by SNaPshot Assay • Microsatellite Genotyping • Golden Gate Assays and Arrays
 Gene Expression on Real Time PCR • Gene expression on Agilent / Microarray / Affymetix • Library construction



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